

MAOI		ImAA	MelMAA (pmol/ml blood)	MeHis	His	Rate of Absorption* (nmol/min)	n
^[14C] -Histamine (10 μ Ci with 82 μ mol/kg)							
	PV	16,422	6,509	1,716	2,179	550	2
	CMA	8,783	4,901	616	559		
	PV	5,173	6,982	9,611	55,763	6,100	2
	CMA	2,030	4,008	4,170	4,606		
Mebanazine (120 μ mol/kg)	PV	1,396	2,319	6,941	11,198	685	4
	CMA	717	1,569	5,570	3,408		
Nialamide (80 μ mol/kg)	PV	3,962	5,741	6,909	8,401	350	3
	CMA	2,995	2,916	9,882	4,460		
Tranlycypromine (14 μ mol/kg)	PV	4,025	2,768	2,545	3,407	700	2
	CMA	1,050	1,254	1,650	810		
Deprenyl (4.5 μ mol/kg)	PV	8,201	3,470	9,733	17,076	510	3
	CMA	6,251	3,309	5,504	4,514		
Clorgyline (24.5 μ mol/kg)	PV	14,420	10,473	10,571	27,650	3,100	3
	CMA	8,439	8,038	6,154	4,769		
Tranlycypromine (80 μ mol/kg)	PV	878	1,661	3,756	10,466	225	2
	CMA	874	1,561	3,948	789		

Values for histamine and metabolites are mean results of serial determinations (approx. 15 per experiment) from n experiments. n = No. of expts.

*The mean absorption rate of [¹⁴C]-compounds at 35 min.

ImAA, Imidazoleacetic acid; MelMAA, *t*-Methylimidazoleacetic acid; MeHis, *t*-Methylhistamine; His, Histamine.

References

- BLACKWELL, B. & MARLEY, E. (1966). Interactions of yeast extracts and their constituents with monoamine oxidase inhibitors. *Br. J. Pharmac. Chemother.*, **26**, 142–161.
- BURKARD, W.P., GEY, K.F. & PLETSCHER, A. (1962). Differentiation of monoamine oxidase and diamine oxidase. *Biochem. Pharmac.* **11**, 177–182.
- IMRIE, P.R., MARLEY, E. & THOMAS, D.V. (1978). Metabolism and distribution of exogenous histamine in cats. *Br. J. Pharmac.* **64**, 109–122.
- ROBINSON, D.S., LOVENBERG, W., KEISER, H. & SJOERDSMA, A. (1968). Effect of drugs on human blood platelet and plasma amine oxidase activity *in vitro* and *in vivo*. *Biochem. Pharmac.* **17**, 109–119.
- SCHAYER, R.W. (1956). The metabolism of histamine in various species. *Br. J. Pharmac.* **11**, 472–473.
- THOMAS, D.V. & MARLEY, E. (1978). Separation of radioactive histamine and some of its metabolites by one dimensional paper chromatography. *J. Chromatog.* **148**, 477–483.

Hepatic microsomal oxidative N-demethylation in rats with renal failure

E.M. HOGAN, P.J. NICHOLLS & A. YOOSUF

Welsh School of Pharmacy, UWIST, Cardiff CF1 3NU, U.K.

The incidence of adverse reactions to drugs is relatively high in patients with chronic renal failure (Smith, Seidl & Chuff, 1966). For some drugs it is possible that this may be related to a decrease in their metabolism (Reidenberg, 1975). In view of the importance of oxidative pathways for drug transformation in the liver, the hepatic microsomal N-demethylation of aminopyrine and ethyl morphine was examined in rats with renal failure.

The five-sixths nephrectomy described by McCance & Morrison (1956) was used to induce renal failure in male Wistar rats (180g). The animals were matched

with pair-fed sham-operated control rats. At 7 and 14 days after nephrectomy, the activities of aminopyrine – (La Du, Gaudette, Trousof & Brodie, 1955) and ethyl morphine – (Holtzman, Gram, Gigon & Gillette, 1968) N-demethylases were determined in the 10,000g supernatant of livers from rats in each set. Hepatic microsomal cytochrome P₄₅₀ was determined by the method of Omura & Sato (1964). Microsomal protein was determined on the 100,000g pellet of the liver-homogenates.

Plasma urea concentrations were significantly raised in the nephrectomized rats at both time intervals but there were no significant differences in body weight (Table 1) or in liver to body weight ratios (overall mean $0.033 \pm .001$) between test and control animals. The Km values for aminopyrine and ethyl morphine demethylation by the hepatic microsomes were unaltered by nephrectomy. However, for the nephrectomized rats at day 14, significant decreases were observed in the rates (V_{max}) of N-demethylation of the two substrates and in the amount of hepatic

cytochrome P₄₅₀ when these values were expressed on the basis of liver weight (Table 1). When the parameters were expressed on the basis of microsomal protein, the differences in mean values between control and test animals were not significant ($P>0.05$). As there was a significant decrease in the concentration of microsomal protein in the liver of nephrectomized rats on day 14 (Table 1), it is likely that this was an important contributor to the decreased oxidative demethylation activity. Similar changes in oxidative metabolism pathways have been found in rats with severe acute renal failure (Leber & Schütterle, 1972).

It is suggested that altered drug metabolism may need to be considered when pharmacokinetic models are developed for application in cases of renal failure.

P.J.N. is in receipt of a grant from the National Kidney Research Fund.

References

- HOLTZMAN, J.L., GRAM, T.E., GIGON, P.L. & GILLETTE, J.R. (1968). The distribution of the components of mixed-function oxidases between the rough and the smooth endoplasmic reticulum of liver cells. *Biochem. J.*, **110**, 407–412.
- LA DU, B.N., GAUDETTE, L., TROUSOF, N. & BRODIE, B.B. (1955). Enzymatic dealkylation of aminopyrine (pyramidon) and other alkylamines. *J. biol. Chem.*, **214**, 741–752.
- LEBER, H.W. & SCHÜTTERLE, G. (1972). Oxidative drug metabolism in liver microsomes from uremic rats. *Kidney Int.*, **2**, 152–158.
- MCCANCE, R.A. & MORRISON, A.B. (1956). The effects of equal and limited rations of water, and of 1, 2 and 3 per cent solutions of sodium chloride on partially nephrectomized and normal rats. *Quart. J. exp. Physiol.*, **41**, 365–386.
- OMURA, T. & SATO, R. (1964). The carbon monoxide-binding pigment of liver microsomes. I. Evidence for its haemoprotein nature. *J. biol. Chem.*, **239**, 2370–2378.
- REIDENBERG, M.M. (1975). Drug metabolism in uraemia. *Clin. Nephrol.*, **4**, 83–85.
- SMITH, J.W., SEIDL, L.G. & CHUFF, L.E. (1966). Studies on the epidemiology of adverse drug reactions. V. Clinical factors influencing susceptibility. *Ann. intern. Med.*, **65**, 629–640.

Table 1 Effects of partial nephrectomy on hepatic N-demethylation of aminopyrine and ethyl morphine in male rats

	Days after nephrectomy			
	7		14	
	C	N	C	N
Body weight (g)	189 ± 5	192 ± 5	215 ± 11	221 ± 8
Plasma urea (mg/100 ml)	40.5 ± 3.3	61.5 ± 3.1	49.0 ± 7.9	70.2 ± 3.1
Cytochrome P ₄₅₀ (μmol/g liver)	27.0 ± 3.3	18.8 ± 2.2	33.2 ± 1.9	22.2 ± 1.7*
(μmol/mg microsomal protein)	0.57 ± 0.04	0.48 ± 0.04	0.55 ± 0.04	0.51 ± 0.03
V _{max} of aminopyrine demethylation (μmol HCHO produced h ⁻¹ g ⁻¹ liver)	4.00 ± 0.36	2.73 ± 0.46	3.18 ± 0.54	1.63 ± 0.28*
(μmol HCHO produced h ⁻¹ mg ⁻¹ microsomal protein)	0.070 ± 0.010	0.060 ± 0.010	0.060 ± 0.014	0.036 ± 0.006
V _{max} of ethyl morphine demethylation (μmol HCHO produced h ⁻¹ g ⁻¹ liver)	12.5 ± 4.3	9.9 ± 0.4	8.5 ± 1.1	3.0 ± 0.6*
(μmol HCHO produced h ⁻¹ mg ⁻¹ microsomal protein)	0.23 ± 0.09	0.22 ± 0.01	0.15 ± 0.03	0.07 ± 0.02
Microsomal protein (mg/g liver)	59.7 ± 8.8	38.9 ± 4.2	57.6 ± 5.2	43.0 ± 3.0*

Values are means ± s.e. means of 5 experiments

C = control N = nephrectomized rats

† 10,000 g supernatant of liver

* $p<0.05$